

# Molecular aspects of gastric cancer development: a mini review

Tarun Sahu

<sup>a</sup>Department of Physiology, All India Institute of Medical Sciences, Raipur, Chhatisgarh, India

\*Corresponding author e-mail: [tarun.sahu50@gmail.com](mailto:tarun.sahu50@gmail.com), [tarunsahu@aiimsraipur.edu.in](mailto:tarunsahu@aiimsraipur.edu.in)

## ABSTRACT

Gastric cancer is still a common problem and is responsible for a large proportion of worldwide cancer mortality, linked to low survival rates. Gastric cancer is a complex, multifaceted process caused by a complicated interaction of environmental and genetic variables. Despite the fact that numerous studies have been conducted on genetic alterations that lead to its genesis, the exact causes remain unknown. Gastric cancer is thought to be caused by various genetic or epigenetic changes, including microsatellite instability (MSI), tumor suppressor genes deactivation, and oncogene overexpression. Apart from that, molecular determinants like cell-cycle regulators and cell adhesion molecules are also responsible for the progression of gastric malignancies. Identifying the etiology of gastric cancer through molecular genetic alterations may assist in the creation of new biomarkers for vulnerability assessment, diagnostic, prognostic, and therapeutic approaches. This review summarizes our current understanding of the molecular basis of gastric cancer, including numerous molecular changes in its pathophysiology and tumorigenesis.

**KEYWORDS:** Gastric cancer, genomic mechanisms, oncogenes, tumor suppressor genes

**Citation:** Sahu. Molecular aspects of gastric cancer development: a mini review. *Polymorphism* 2022; 8: 26-36.

Received: September 11, 2021; revised: October 12, 2021; Accepted: October 15, 2021

## INTRODUCTION

Gastric cancer is still the second biggest reason of death worldwide among all cancers. Although death rates have been decreasing, most patients who develop GC are aged 65 to 74. The disease affects men twice as often as women. African Americans, Spanish Americans, and Native Americans are more likely to develop GC than white Americans (Koh and Wang 2001). In Indian subcontinent, the rate of gastric cancer in the north-eastern areas are greater than the rest of the country. According to studies, the rate of gastric cancer in the urban regions are 3.0–13.2 when compared to global rate (4.1–95.5), and these are high among males (50.6) than females (23.3) (Dikshit et al. 2011; Sharma and Radhakrishnan 2011). According to Cancer Statistics, 2020, gastric cancer is projected among five most common cancers in 2020 for males, which accounts for 36% out of all cancers (Mathur et al. 2020).

Gastric cancer is a complex, multifaceted phenomenon caused by a complicated interaction of environmental and genetic variables. Apart from genetic factors, the non-genetic determinants of gastric cancer consist of poor dietary habits, *H. pylori* infection, family history, and tobacco smoking (Fujino et al. 2002; Lazarevic et al. 2010). Diet has a significant influence in the development of gastric cancer. High salt intake has been linked to an increase in gastric cancer morbidity and death. High-salt foods may harm the stomach mucosa, resulting in gastritis, excessive DNA synthesis, and increased cell replication. Chronic salt consumption may exacerbate *H. pylori*-related carcinogenesis by promoting *H. pylori* proliferation, colonisation, and mucosal atrophy (Furihata, Ohta, and Katsuyama 1996; X.-Q. Wang, Terry, and Yan 2009). For decades, researchers have been aware of the impact that alcohol intake plays a crucial role in carcinogenesis. The ethanol in alcoholic beverages causes oxidative stress, which damages DNA and impairs its repair. Acetaldehyde is the first

metabolite generated during alcohol breakdown and it may have a more prominent role in the carcinogenic effect of ethanol on the mucosa because of its numerous mutagenic effects on DNA (Baan et al. 2007; Salaspuro 2011).

Obesity is another major concern, which raises the risk of gastric malignancies. Obesity is thought to be responsible for 14% of all cancer deaths in men and 20% in women (Calle et al. 2003). Obesity-related gastro-oesophageal reflux, insulin resistance, abnormal levels of adiponectin, leptin, and ghrelin, as well as an unusually high blood level of insulin-like growth factor, are all possible pathways connecting obesity to gastric cancer (Li et al. 2012; Donohoe et al. 2014; Karczewski et al. 2019). The gut microbiome/ microbiota are varied population of bacteria that lives in the human gut. Human gut bacteria are also thought to have a key role in the genesis of gastric malignancies, particularly by dysbiosis (imbalance in the gut microbiota, which can contribute to a variety of clinical disorders, especially GC) (Thursby and Juge 2017). The entire research till date on the gastric microbiota performed shows that the microbiota changes as the gastric mucosa progresses from normal to cancerous. But, these studies have shown conflicting results when it comes to whether the microbiota of stomach cancer patients has more or less variety than the microbiota of healthy people; therefore, there is a scarcity of evidence on the relationship between the gut microbiota and GC occurrence (Bik et al. 2006; Dicksved et al. 2009; Aviles-Jimenez et al. 2014; L. Wang et al. 2016; Stewart, Wu, and Chen 2020).

However, the exact process of GC formation is yet unknown. Metabolic pathway metabolites play a critical role in controlling cell differentiation during carcinogenesis as well as normal growth. A recent study shows that glutamine metabolism plays a crucial role in tumorigenesis because it is an excellent source of reduced nitrogen for the synthesis of purine and pyrimidine bases, as well as

proteins(Addeo et al. 2021; Ratre et al. 2021). Despite many advancements in the treatment strategies of GC, the tumor's resistance mechanisms provide a significant barrier to the development of successful cancer treatments. Recent data suggest that miRNAs have a critical role in tumor development and also cause chemoresistance in GC. The expression of certain kinds of microRNAs is increased in GC. Thus it has potential diagnostic values in the prediction of the disease(Verma et al. 2020; Verma and Bhaskar 2020). Although there

has been significant improvement in the diagnostic and therapeutic approach, the life expectancy remains static and low, with approximately 20% of individuals with GC surviving for five years(Singh et al., n.d.). Due to this, a comprehensive understanding of the genetic basis of GC is required to develop innovative GC preventive and treatment techniques. The objective of this review is to emphasize the pathophysiology of the most common forms of gastric cancer.

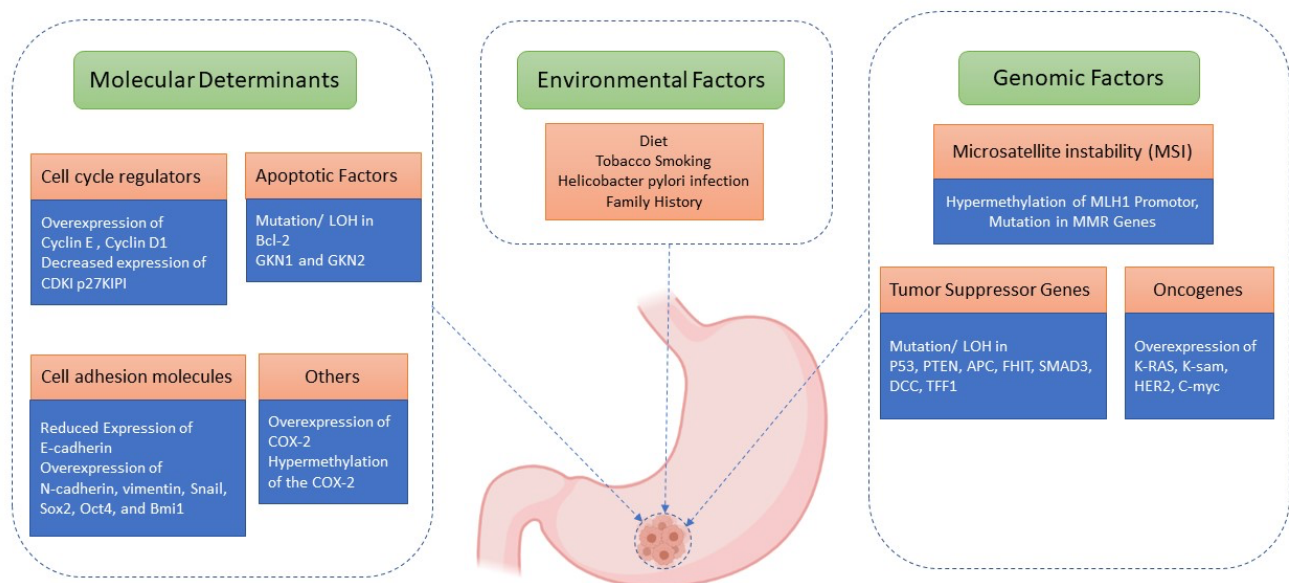


Fig. 1. Factors that cause the development of gastric cancer

Gastric carcinoma, which comprises of more than 50% of all gastric cancers, is classified into two subgroups by Lauren's classification: intestinal and diffuse(LAUREN 1965). The intestinal form is defined by cancerous epithelial cells with cohesion and glandular development that infiltrate the surrounding regions. It is linked with *H. pylori* infection, overweight, and other dietetic aspects like a huge salt consumption, smoked meats, and various food preservatives. The diffuse category is defined by cancer cells with low development and non-cohesiveness. It is also known to be developed by *H. pylori* infection with chronic inflammation(Cristescu et al. 2015; Crew and

Neugut 2006). Factors that cause the development of gastric cancer are depicted in Figure 1.

### Genomic mechanisms of gastric carcinogenesis

Although gastric cancer's genomic mechanisms are complex and poorly understood, rapid developments in cancer biology have shown that converting a normal epithelial cell into a cancerous cell is a sequential process involving the collection of numerous genetic alterations (Chan et al. 1999; Correa 1992; Powell et al. 1992). Gastric cancer is thought to be caused by multiple genetic and epigenetic changes, as genomic instability is an essential discovery in gastric cancer(Hanahan and

Weinberg 2000). Many molecular anomalies/genomic instability were documented, including microsatellite instability (MSI), deactivation of tumor suppressor genes, and overexpression of oncogenes (Correa 1992; Werner et al. 2001; Meining et al. 2001).

### Microsatellite instability (MSI)

MSI is associated with replication errors in microsatellite sequences due to a deficiency in the mismatch repair system. A set of mismatch repair (MMR) proteins detects and corrects these errors. MSI is caused by the deactivation or lack of MMR genes, typically MLH1 or MSH2, which often cause other genetic alterations, such as down-regulation of the tumor suppressor genes (Hudler 2012; Buermeyer et al. 1999). In MSI, the number of microsatellite repeats in tumor cells is different from normal cells. MSI is among the early changes in tumorigenesis that lead to genomic instability (Miyoshi et al. 2001). Rather than searching for a specific gene, the MSI method can be utilized as a screening tool for early cancer detection. CpG island methylator pathway defines the methylation in gastric cancer. Hypermethylation in the hMLH1 gene is linked with functional inactivation or silencing of the gene. Epigenetic hypermethylation of the MLH1 promoter has been associated with the formation of more than half of MSI-H-positive gastric malignancies.

In contrast, alterations in MLH1 and MSH2 have been detected in 12 to 15 percent of MSI-H-positive gastric carcinomas (Toyota et al. 1999; Leung et al. 1999; Hudler et al. 2004; Etoh et al. 2004). BCL2 associated X gene, MutS homolog 3 and 6, E2F Transcription Factor 4, TGFRII, insulin-like growth factor receptor II, TCF 4, RIZ, CASPASE 5, FAS, BCL10, and APAF1 were reported to be mutated in gastric cancers with MSI. These genes have been linked to the control of cell-cycle transition and apoptosis (Fiocca et al. 2001; El-Rifai and Powell 2002; Ottini et al. 2006).

### Tumor suppressor genes

Tumor suppressor genes prevent cells from turning malignant by repairing DNA, which inhibits cell growth and triggers apoptosis. Tumor suppressor genes have functions in various cell activities such as cell adhesion, cell interaction, cytoplasmic signaling processes, and nuclear transcription. Tumor suppressor gene inactivation because of mutations and/or loss of heterozygosity (LOH) is another common occurrence in gastric carcinogenesis. Mutation or loss of function of the p53 gene was observed in 80% of gastric carcinomas (Werner et al. 2001; Shiao et al. 1994), and changes in the p53s function caused by LOH include a high frequency of p53 alterations and DNA methylation in rare cases (Verma, Falco, and Bhaskar 2020). The p53 gene is known to be the primary regulator of cell division. p53's biological purpose is to protect genomic integrity, and cells that lack normal p53 function will not initiate apoptosis and hence do not control the growth of tumors (Vousden and Prives 2005). Over 50% of malignancies, comprising leukemia, breast, colon, and lung carcinoma, have abnormal p53 expression, and p53 mutation perhaps has the highest prevalence in all of them. p53 is a crucial player in the cell cycle; after DNA damage, p53 inhibits the G1/S shift, thus enabling DNA repair and apoptosis. LOH of the chromosome 17p, which codes p53, is seen in 62% of the atypical intestinal forms of GCs developed by gastric carcinoma (H. S. Kim et al. 2001). PTEN is another tumor suppressor gene that was discovered in 1977. It controls multiplication, apoptosis, motility, adherence, and genomic integrity through a sophisticated network structure that governs a range of cellular activities and signaling pathways. It is a lipid phosphatase that inhibits the PI3K signaling pathway by obstructing PIP3 dependent processes like activation of AKT (Xi and Chen 2015). So, upon the loss of expression of PTEN, the PIP3 level will be increased, which also increases AKT activation, which ultimately inhibits apoptosis (Xu, Yang, and Lu 2014). PTEN protein expression is substantially lower

in gastric cancer in contrast to normal. Genes of phosphatidylinositol 3-kinases, Protein kinase B, Matrix Metalloproteinase 2 and 9, and nuclear factor-B (NF-Bp65) protein are, on the other hand, overexpressed in GC(Wen et al. 2010). Alterations or LOH of the APC (tumor suppressor gene) are also found in sixty percent of intestinal-form of gastric tumors and approximately twenty five percent of adenomas(Horii et al. 1992; Wright and Williams 1993; Tahara 1995). According to a recent study, somatic mutation of the APC gene has a significant role in the etiology of benign gastric tumors, but only a minor impact in its development to malignant carcinoma(J.-H. Lee et al. 2002).

Further, the APC protein plays a vital role in the degradation of  $\beta$ -catenin, a transcription factor of the Wnt signaling pathway(Hsieh and Huang 1995; Caca et al. 1999; Park et al. 1999). APC protein binds specifically to  $\beta$ -catenin, and after combining with transcription factor LEF-1,  $\beta$ -catenin move into the nucleus, at which it controls gene expression. Mutation in  $\beta$ -catenin/lymphoid-enhancing factor have been discovered in intestinal-type of GC (Park et al. 1999; Peifer 1999). In patients with familial adenomatous polyposis (FAP), where 60–80 % of APC genes are mutated, there is also evidence of a link between APC and gastric cancer (Kinzler et al. 1991; Lynch and Lynch 1998), and individuals with FAP have ten times increased lifetime risk of gastric cancer than ordinary people(Zwick et al. 1997). APC is also altered by extensive methylation and leads to premalignant stages leading to gastric cancer. Methylation in APC has been found in greater than 75% of the intestinal forms of GC(Esteller et al. 2000; Sarbia et al. 2004). p16 is a CDK inhibitor that arrests the cell cycle at the G1 phase, and the reduced expression of p16 is seen in many intestinal GCs. This reduced expression of p16 is due to hypermethylation(Sarbia et al. 2004). FHIT is another tumor suppressor gene from FRA3B at the 3p14.2 site, which was abnormally expressed due to genetic alterations in most gastric cancers(Baffa et al. 1998; Ohta et al. 1996). SMAD3 is another tumor suppressor whose absence is linked to a severe

phase and a worse prognosis in individuals with gastric cancer. It is more prevalent in the intestinal form than that in the diffuse form. DCC is a type of tumor suppressor gene located on chromosome 18q. One study found reduced mRNA expression of DCC in 52 cases of gastric malignancies and stated that the reduced expression was strictly related to liver metastasis(Yoshida et al. 1998). Trefoil factor family 1 (TFF1) is a gene found on chromosome 21q22, and its synthesis occurs in the stomach mucosa and gastrointestinal cells of injured tissue. This gene is also found deleted in most gastric cancers in LOH studies, and in some cases, its altered expression due to mutation is also seen(Bossenmeyer-Pourié et al. 2002; Park et al. 2000).

### Oncogenes

An oncogene is a mutated gene responsible for cancer progression, and these genes are expressed at high levels in cancer cells. RAS are oncogenes that are not found in regular cells but are triggered in cancer cells due to massive mutations that occur throughout tumor growth. K-ras is a cellular signaling protein that plays a significant role in developing pancreatic and colon cancers(Pellegata et al. 1992; Soh et al. 1993). K-sam, a member of the FGFR, is also shown to be overexpressed in diffuse-forms of gastric tumors(Kato 2003). The c-erbB-2 OR HER-2/neu is a TKI receptor-like EGFR 2, HER2, and EGFR or HER1, and it is also overexpressed in all types of GCs(García et al. 2003). In intestinal-type GC, serum c-erbB-2 levels are considerably elevated in comparison with diffuse GC(Vizoso et al. 2004). The majority of research on this protein has found that HER-2/neu status has a substantial predictive value, and HER-2/neu overexpression was identified as a possible biomarker of prognosis in gastric malignancies(Mizutani et al. 1993; Allgayer et al. 2000; Y.-L. Wang et al. 2002). C-myc is an oncogene found on chromosome 8 that expresses a nuclear phosphoprotein that functions as a transcription factor (TF) by stimulating and inhibiting the expression of target genes. It also

involves regulating many genes responsible for critical cellular processes such as multiplication, development, differentiation, angiogenesis, DNA repair, and cell death. If myc gene is unregulated, it can lead to tumor development (Battey et al. 1983). C-myc overexpression is observed in more than 40 percent of gastric cancers. This is an initial phase in the development of both intestinal as well as diffuse gastric cancers. However, it is significantly more prevalent in the intestinal form as compared to the diffuse form. The C-myc gene has been reported to be highly expressed in benign forms such as chronic atrophic gastritis, gastric ulcer, and *H. pylori* infection (Calcagno et al. 2008; Dang et al. 2006).

## Molecular determinants of gastric cancer

### Cell cycle regulators and apoptosis

Cell cycle regulators such as cyclin proteins control the cell cycle by checkpoints by attaching and triggering particular cyclin-dependent kinases. G1-S phase progression is controlled via cyclin D, E, and A, and its kinases like CDK 2,4, and 6, whereas G2/M shift is controlled via B-type cyclin-associated kinase. Cell-cycle regulator abnormalities have also been associated with gastric cancer formation and development through uncontrolled cell growth (Yasui et al. 2001). Overexpression of cyclin E was associated with later cancer stage and metastatic spread in 10% of diffuse-form gastric cancers and 20% of intestinal-form gastric cancers (Yasui et al. 1997). Moreover, overexpression of cyclin D1 was identified in about 50% of gastric carcinomas, with the intestinal form being more common than the diffuse form (Müller et al. 1999), and the reduced expression of CDKI p27KIP1 in GC is usually linked to the degree of tumor progression and the occurrence of lymph node metastasis (Yasui et al. 1997).

Apoptosis, or programmed cell death, plays a vital role in regulating a cell's entity. FAS, TNF, and bcl-2, are apoptosis-regulating genes that play a role in cancer development. Bcl-2 was discovered in a human leukemia line at the chromosomal

breakpoint t (14;18) (Tsujiimoto et al. 1984; 1985). Around fifteen Bcl-2 family member proteins in mammalian cells contained proteins that trigger and inhibit apoptosis (Gross, McDonnell, and Korsmeyer 1999). Although LOH at the bcl-2 region has been linked with intestinal-form of gastric carcinoma, the expression of an apoptotic receptor antigen identified by SC-1 antibody is predominantly found in diffuse-form of cancers (Werner et al. 2001; Vollmers et al. 1997; Klein, Vollmers, and MullerHermelink 1996). GKN1 has been found to block the G2/M cell cycle, inhibiting tumor cell development and lowering the number of cell colonies (Yan et al. 2011). In vitro studies revealed that both GKN1 and GKN2 had been found in the gastric mucosal lining of healthy people and the gastric mucosa of patients with GC; however, there is a contrast to those by superficial gastritis reduction in GKN1 mRNA expression and protein (Yoshihara et al. 2006; Oien et al. 2004; X.-Y. Guo et al. 2014; Zhang et al. 2010; Yan et al. 2011).

### Cell adhesion molecules

Gastric cancer also appears to be linked to changes in adhesion molecules. Cadherins like E, N, P, R, and M are predominant, with their names derived from the type of cell in which expression was initially identified (Shimada et al. 2012). E-cadherin is a  $\beta$ -catenin binding partner that is important for intercellular adhesion and epithelial tissue structural integrity. E-cadherin is a member of the cadherin superfamily and is engaged in epithelial phenotype maintenance. E-cadherin expression is often diminished or lost in gastric malignancies, most likely due to hypermethylation of the E-cadherin promoter (Tamura et al. 2000; Mingchao et al. 2001). Somatic changes in E-cadherin were found in 50% of patients with diffuse-form of gastric malignancy. In contrast, deletion of the leftover alleles, leading to full protein deactivation, was reported in more than 75% of cases with mutation (Becker et al. 1994; Oda et al. 1994). Oxygen deprivation is a strong activator of GC EMT (epithelial-mesenchymal transition). E-cadherin levels drop in hypoxic

conditions while N-cadherin, vimentin, Snail, Sox2, Oct4, and Bmi1 levels rise, suggesting that the hypoxia promotes epithelial-mesenchymal transition and cytoskeleton remodeling (J. Guo et al. 2016).

### Cyclooxygenase (COX)-2

Nonsteroidal anti-inflammatory medications block COX-2, the primary enzyme involved in converting arachidonic acid to prostanooids. COX-2 overexpression has been related to inflammatory processes and carcinogenesis in various human malignancies, most notably colorectal, esophageal, and gastric cancers (B P van Rees and Ristimäki 2001; Rajnakova et al. 2001; Bastiaan P van Rees et al. 2002). Overexpression of COX-2 is typical in the intestinal form of gastric cancer, and dysplastic precursor lesions, representing that COX-2 expression is involved in gastric carcinogenesis at an early stage (Lim et al. 2000; Saukkonen et al. 2001). Furthermore, in gastric cancer, COX-2 overexpression is significantly and inversely associated with MSI (T. L. Lee et al. 2001). Hypermethylation of the COX-2 CpG island in gastric cancer cell lines has been demonstrated to produce COX-2 transcriptional suppression. Furthermore, COX-2 expression in gastric epithelial cells is controlled by *H. pylori*-stimulated promoter methylation (H. Kim, Lim, and Kim 2001; Song et al. 2001).

### CONCLUSION

Several important discoveries about the molecular pathogenesis of GC have been made since the last decade. These findings may enhance our knowledge of molecular interactions involved in gastric carcinogenesis and ultimately, lead to the discovery of novel treatment regimens that precisely target genetic alterations for this common disease. The characterization of GC at the molecular and genetic level supports the idea that this disease is very divergent. As evidenced by the complexity and variety of gastric tumors, gastric cancer is the result of a cascade of events involving diverse

genetic and epigenetic alterations in multiple genes in combination with the host's genetic lineage and external conditions. Further research and meta-analyses are required to develop panels of biomarkers for assessment of vulnerability, diagnostic, prognostic, and therapeutic strategy.

### Acknowledgments

The authors would like to thank the parent institution for support.

### Author's contribution

TS designed, planned, reviewed and prepared the manuscript.

### Conflict of interest

The author has no conflict of interest.

### Source of Funding

The author declares that this research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

### Declaration of originality

The author has declared that the data/text presented in this manuscript is original and no text, figure or data has been copied from any other source without appropriate citation.

### Jurisdiction and maps

Polymorphism and Peer Publishers remain neutral to the jurisdictional claims, maps, boundaries and institutional affiliations shown or claimed in any of the articles published.

### REFERENCES

- Koh T, Wang T. Tumors of the stomach. In: in: Feldman M., editor. Sleisenger and Fordtran's gastrointestinal and liver disease. 7th ed. Saunders, Philadelphia; 2001. p. 829–51.
- Dikshit RP, Mathur G, Mhatre S, Yeole BB. Epidemiological review of gastric cancer in India. Indian J Med Paediatr Oncol Off J Indian Soc Med Paediatr Oncol. 2011

- Jan;32(1):3–11.
- Sharma A, Radhakrishnan V. Gastric cancer in India. *Indian J Med Paediatr Oncol Off J Indian Soc Med Paediatr Oncol*. 2011 Jan;32(1):12–6.
- Mathur P, Sathishkumar K, Chaturvedi M, Das P, Sudarshan KL, Santhappan S, et al. Cancer Statistics, 2020: Report From National Cancer Registry Programme, India. *JCO Glob Oncol*. 2020;(6):1063–75.
- Fujino Y, Tamakoshi A, Ohno Y, Mizoue T, Tokui N, Yoshimura T. Prospective study of educational background and stomach cancer in Japan. *Prev Med (Baltim)*. 2002 Aug;35(2):121–7.
- Lazarevic K, Nagorni A, Rancic N, Milutinovic S, Stosic L, Ilijev I. Dietary factors and gastric cancer risk: hospital-based case control study. *J BUON*. 2010;15(1):89–93.
- Furihata C, Ohta H, Katsuyama T. Cause and effect between concentration-dependent tissue damage and temporary cell proliferation in rat stomach mucosa by NaCl, a stomach tumor promoter. *Carcinogenesis*. 1996;17(3):401–6.
- Wang X-Q, Terry P-D, Yan H. Review of salt consumption and stomach cancer risk: epidemiological and biological evidence. *World J Gastroenterol*. 2009 May;15(18):2204–13.
- Baan R, Straif K, Grosse Y, Secretan B, El Ghissassi F, Bouvard V, et al. Carcinogenicity of alcoholic beverages. *Lancet Oncol*. 2007 Apr;8(4):292–3.
- Salaspuro M. Acetaldehyde and gastric cancer. *J Dig Dis*. 2011 Apr;12(2):51–9.
- Calle EE, Rodriguez C, Walker-Thurmond K, Thun MJ. Overweight, obesity, and mortality from cancer in a prospectively studied cohort of U.S. adults. *N Engl J Med*. 2003 Apr;348(17):1625–38.
- Li Q, Zhang J, Zhou Y, Qiao L. Obesity and gastric cancer. *Front Biosci (Landmark Ed)*. 2012 Jun;17:2383–90.
- Donohoe CL, O'Farrell NJ, Doyle SL, Reynolds J V. The role of obesity in gastrointestinal cancer: evidence and opinion. *Therap Adv Gastroenterol*. 2014 Jan;7(1):38–50.
- Karczewski J, Begier-Krasińska B, Staszewski R, Popławska E, Gulczynska-Elhadi K, Dobrowolska A. Obesity and the Risk of Gastrointestinal Cancers. *Dig Dis Sci*. 2019;64(10):2740–9.
- Thursby E, Juge N. Introduction to the human gut microbiota. *Biochem J*. 2017 May 16;474(11):1823–36.
- Bik EM, Eckburg PB, Gill SR, Nelson KE, Purdom EA, Francois F, et al. Molecular analysis of the bacterial microbiota in the human stomach. *Proc Natl Acad Sci U S A*. 2006 Jan;103(3):732–7.
- Dicksved J, Lindberg M, Rosenquist M, Enroth H, Jansson JK, Engstrand L. Molecular characterization of the stomach microbiota in patients with gastric cancer and in controls. *J Med Microbiol*. 2009 Apr;58(Pt 4):509–16.
- Aviles-Jimenez F, Vazquez-Jimenez F, Medrano-Guzman R, Mantilla A, Torres J. Stomach microbiota composition varies between patients with non-atrophic gastritis and patients with intestinal type of gastric cancer. *Sci Rep*. 2014 Feb;4:4202.
- Wang L, Zhou J, Xin Y, Geng C, Tian Z, Yu X, et al. Bacterial overgrowth and diversification of microbiota in gastric cancer. *Eur J Gastroenterol Hepatol*. 2016 Mar;28(3):261–6.
- Stewart OA, Wu F, Chen Y. The role of gastric microbiota in gastric cancer. *Gut Microbes*. 2020;11(5):1220–30.
- Addeo M, Di Paola G, Verma HK, Laurino S, Russi S, Zoppoli P, et al. Gastric Cancer Stem Cells: A Glimpse on Metabolic Reprogramming. *Front Oncol*. 2021;11:2308.
- Ratre YK, Verma HK, Mehta A, Soni VK, Sonkar SC, Shukla D, et al. Therapeutic Targeting of Glutamine Metabolism in Colorectal Cancer BT - Colon Cancer Diagnosis and Therapy: Volume 2. In: Vishvakarma NK, Nagaraju GP, Shukla D, editors. Cham: Springer International Publishing; 2021. p. 333–56.
- Verma HK, Ratre YK, Mazzone P, Laurino S, Bhaskar LVKS. Micro RNA facilitated chemoresistance in gastric cancer: a novel biomarkers and potential therapeutics. *Alexandria J Med*. 2020;56(1):81–92.
- Verma HK, Bhaskar L. MicroRNA a small magic bullet for gastric cancer. *Gene*. 2020 Aug;753:144801.
- Singh J, Kumar P, Verma K, Tiwary SK, Narayan G, Dixit VK. Molecular genetic changes in gastric carcinoma. *Int J Mol Immuno Oncol*. 6.
- Lauren P. The two histological main types of gastric carcinoma: diffuse and so-called intestinal-type carcinoma. An attempt at a histo-clinical classification. *Acta Pathol Microbiol Scand*. 1965;64:31–49.
- Cristescu R, Lee J, Nebozhyn M, Kim K-M, Ting JC, Wong SS, et al. Molecular analysis of gastric cancer identifies subtypes associated with distinct clinical outcomes. *Nat Med*. 2015 May;21(5):449–56.
- Crew KD, Neugut AI. Epidemiology of gastric cancer. *World J Gastroenterol*. 2006 Jan;12(3):354–62.
- Chan AO, Luk JM, Hui WM, Lam SK. Molecular biology of gastric carcinoma: from laboratory to bedside. *J Gastroenterol Hepatol*. 1999 Dec;14(12):1150–60.
- Correa P. Human gastric carcinogenesis: a multistep and multifactorial process--First American Cancer Society Award Lecture on Cancer Epidemiology and Prevention. *Cancer Res*. 1992 Dec;52(24):6735–40.
- Powell SM, Zilz N, Beazer-Barclay Y, Bryan TM, Hamilton SR, Thibodeau SN, et al. APC mutations occur early during colorectal tumorigenesis. *Nature*. 1992 Sep;359(6392):235–7.
- Hanahan D, Weinberg RA. The Hallmarks of Cancer. *Cell*. 2000 Jan 7;100(1):57–70.
- Werner M, Becker KF, Keller G, Höfler H. Gastric adenocarcinoma: pathomorphology and molecular pathology. *J Cancer Res Clin Oncol*. 2001 Apr;127(4):207–16.
- Meining A, Morgner A, Miehke S, Bayerdörffer E, Stolte M. Atrophy-metaplasia-dysplasia-carcinoma sequence in the stomach: a reality or merely an hypothesis? *Best Pract Res Clin Gastroenterol*. 2001 Dec;15(6):983–98.
- Hudler P. Genetic Aspects of Gastric Cancer Instability. Belkhir A, editor. *Sci World J*. 2012;2012:761909.



- Buermeyer AB, Deschênes SM, Baker SM, Liskay RM. Mammalian DNA Mismatch Repair. *Annu Rev Genet.* 1999;33(1):533–64.
- Miyoshi E, Haruma K, Hiyama T, Tanaka S, Yoshihara M, Shimamoto F, et al. Microsatellite instability is a genetic marker for the development of multiple gastric cancers. *Int J cancer.* 2001 Nov;95(6):350–3.
- Toyota M, Ahuja N, Suzuki H, Itoh F, Ohe-Toyota M, Imai K, et al. Aberrant methylation in gastric cancer associated with the CpG island methylator phenotype. *Cancer Res.* 1999 Nov;59(21):5438–42.
- Leung SY, Yuen ST, Chung LP, Chu KM, Chan AS, Ho JC. hMLH1 promoter methylation and lack of hMLH1 expression in sporadic gastric carcinomas with high-frequency microsatellite instability. *Cancer Res.* 1999 Jan;59(1):159–64.
- Hudler P, Vouk K, Vouk K, Liovic M, Repse S, Juvan R, et al. Mutations in the hMLH1 gene in Slovenian patients with gastric carcinoma. *Clin Genet.* 2004;65:405–11.
- Etoh T, Kanai Y, Ushijima S, Nakagawa T, Nakanishi Y, Sasako M, et al. Increased DNA methyltransferase 1 (DNMT1) protein expression correlates significantly with poorer tumor differentiation and frequent DNA hypermethylation of multiple CpG islands in gastric cancers. *Am J Pathol.* 2004 Feb;164(2):689–99.
- Fiocca R, Luinetti O, Villani L, Mastracci L, Quilici P, Grillo F, et al. Molecular mechanisms involved in the pathogenesis of gastric carcinoma: interactions between genetic alterations, cellular phenotype and cancer histotype. *Hepatogastroenterology.* 2001;48(42):1523–30.
- El-Rifai W, Powell SM. Molecular biology of gastric cancer. *Semin Radiat Oncol.* 2002 Apr;12(2):128–40.
- Ottini L, Falchetti M, Lupi R, Rizzolo P, Agnese V, Colucci G, et al. Patterns of genomic instability in gastric cancer: clinical implications and perspectives. *Ann Oncol.* 2006 Jun 1;17:vii97–102.
- Shiao YH, Rugge M, Correa P, Lehmann HP, Scheer WD. p53 alteration in gastric precancerous lesions. *Am J Pathol.* 1994 Mar;144(3):511–7.
- Verma HK, Falco G, Bhaskar LVKS. Molecular Signaling Pathways Involved in Gastric Cancer Chemoresistance. In: Raju GSR, Bhaskar LVKS, editors. *Theranostics Approaches to Gastric and Colon Cancer.* Singapore: Springer Singapore; 2020. p. 117–34.
- Vousden KH, Prives C. P53 and prognosis: new insights and further complexity. *Cell.* 2005 Jan;120(1):7–10.
- Kim HS, Woo DK, Bae SI, Kim YI, Kim WH. Allelotype of the adenoma-carcinoma sequence of the stomach. *Cancer Detect Prev.* 2001;25(3):237–44.
- Xi Y, Chen Y. Oncogenic and Therapeutic Targeting of PTEN Loss in Bone Malignancies. *J Cell Biochem.* 2015 Sep;116(9):1837–47.
- Xu W-T, Yang Z, Lu N-H. Roles of PTEN (Phosphatase and Tensin Homolog) in gastric cancer development and progression. *Asian Pac J Cancer Prev.* 2014;15(1):17–24.
- Wen Y-G, Wang Q, Zhou C-Z, Qiu G-Q, Peng Z-H, Tang H-M. Mutation analysis of tumor suppressor gene PTEN in patients with gastric carcinomas and its impact on PI3K/AKT pathway. *Oncol Rep.* 2010 Jul;24(1):89–95.
- Horii A, Nakatsuru S, Miyoshi Y, Ichii S, Nagase H, Kato Y, et al. The APC gene, responsible for familial adenomatous polyposis, is mutated in human gastric cancer. *Cancer Res.* 1992 Jun;52(11):3231–3.
- Wright PA, Williams GT. Molecular biology and gastric carcinoma. *Gut.* 1993 Feb;34(2):145–7.
- Tahara E. Genetic alterations in human gastrointestinal cancers. The application to molecular diagnosis. *Cancer.* 1995 Mar;75(6 Suppl):1410–7.
- Lee J-H, Abraham SC, Kim H-S, Nam J-H, Choi C, Lee M-C, et al. Inverse relationship between APC gene mutation in gastric adenomas and development of adenocarcinoma. *Am J Pathol.* 2002 Aug;161(2):611–8.
- Hsieh LL, Huang YC. Loss of heterozygosity of APC/MCC gene in differentiated and undifferentiated gastric carcinomas in Taiwan. *Cancer Lett.* 1995 Sep;96(2):169–74.
- Caca K, Kolligs FT, Ji X, Hayes M, Qian J, Yahanda A, et al. Beta- and gamma-catenin mutations, but not E-cadherin inactivation, underlie T-cell factor/lymphoid enhancer factor transcriptional deregulation in gastric and pancreatic cancer. *Cell growth Differ Mol Biol J Am Assoc Cancer Res.* 1999 Jun;10(6):369–76.
- Park WS, Oh RR, Park JY, Lee SH, Shin MS, Kim YS, et al. Frequent somatic mutations of the beta-catenin gene in intestinal-type gastric cancer. *Cancer Res.* 1999 Sep;59(17):4257–60.
- Peifer M. Signal transduction. Neither straight nor narrow. *Vol. 400, Nature.* England; 1999. p. 213–5.
- Kinzler KW, Nilbert MC, Su LK, Vogelstein B, Bryan TM, Levy DB, et al. Identification of FAP locus genes from chromosome 5q21. *Science.* 1991 Aug;253(5020):661–5.
- Lynch HT, Lynch JF. Genetics of colonic cancer. *Digestion.* 1998 Aug;59(5):481–92.
- Zwick A, Munir M, Ryan CK, Gian J, Burt RW, Leppert M, et al. Gastric adenocarcinoma and dysplasia in fundic gland polyps of a patient with attenuated adenomatous polyposis coli. *Gastroenterology.* 1997 Aug;113(2):659–63.
- Esteller M, Sparks A, Toyota M, Sanchez-Cespedes M, Capella G, Peinado MA, et al. Analysis of adenomatous polyposis coli promoter hypermethylation in human cancer. *Cancer Res.* 2000 Aug;60(16):4366–71.
- Sarbia M, Geddert H, Klump B, Kiel S, Iskender E, Gabbert HE. Hypermethylation of tumor suppressor genes (p16INK4A, p14ARF and APC) in adenocarcinomas of the upper gastrointestinal tract. *Int J cancer.* 2004 Aug;111(2):224–8.
- Baffa R, Veronese ML, Santoro R, Mandes B, Palazzo JP, Rugge M, et al. Loss of FHIT expression in gastric carcinoma. *Cancer Res.* 1998 Oct;58(20):4708–14.
- Ohta M, Inoue H, Cotticelli MG, Kastury K, Baffa R, Palazzo J, et al. The FHIT gene, spanning the chromosome 3p14.2 fragile site and renal carcinoma-associated t(3;8) breakpoint, is abnormal in digestive tract cancers. *Cell.* 1996

- Feb;84(4):587–97.
- Yoshida Y, Itoh F, Endo T, Hinoda Y, Imai K. Decreased DCC mRNA expression in human gastric cancers is clinicopathologically significant. *Int J cancer*. 1998 Dec;79(6):634–9.
- Bossemeyer-Pourié C, Kannan R, Ribieras S, Wendling C, Stoll I, Thim L, et al. The trefoil factor 1 participates in gastrointestinal cell differentiation by delaying G1-S phase transition and reducing apoptosis. *J Cell Biol*. 2002 May;157(5):761–70.
- Park WS, Oh RR, Park JY, Lee JH, Shin MS, Kim HS, et al. Somatic mutations of the trefoil factor family 1 gene in gastric cancer. *Gastroenterology*. 2000 Sep;119(3):691–8.
- Pellegata NS, Losekoot M, Fodde R, Pugliese V, Saccomanno S, Renault B, et al. Detection of K-ras mutations by denaturing gradient gel electrophoresis (DGGE): a study on pancreatic cancer. *Anticancer Res*. 1992;12(5):1731–5.
- Soh K, Yanagisawa A, Hiratsuka H, Sugano H, Kato Y. Variation in K-ras codon 12 point mutation rate with histological atypia within individual colorectal tumors. *Jpn J Cancer Res*. 1993 Apr;84(4):388–93.
- Kato M. WNT2 and human gastrointestinal cancer (review). *Int J Mol Med*. 2003 Nov;12(5):811–6.
- García I, del Casar JM, Corte MD, Allende MT, García-Muñiz JL, Vizoso F. Epidermal growth factor receptor and c-erbB-2 contents in unresectable (UICC R1 or R2) gastric cancer. *Int J Biol Markers*. 2003;18(3):200–6.
- Vizoso FJ, Corte MD, Alvarez A, García I, del Casar JM, Bongera M, et al. Membranous levels of c-erbB-2 oncoprotein in gastric cancer: their relationship with clinicopathological parameters and their prognostic significance. *Int J Biol Markers*. 2004;19(4):268–74.
- Mizutani T, Onda M, Tokunaga A, Yamanaka N, Sugisaki Y. Relationship of C-erbB-2 protein expression and gene amplification to invasion and metastasis in human gastric cancer. *Cancer*. 1993 Oct;72(7):2083–8.
- Allgayer H, Babic R, Gruetzner KU, Tarabichi A, Schildberg FW, Heiss MM. c-erbB-2 is of independent prognostic relevance in gastric cancer and is associated with the expression of tumor-associated protease systems. *J Clin Oncol Off J Am Soc Clin Oncol*. 2000 Jun;18(11):2201–9.
- Wang Y-L, Sheu B-S, Yang H-B, Lin P-W, Chang Y-C. Overexpression of c-erbB-2 proteins in tumor and non-tumor parts of gastric adenocarcinoma--emphasis on its relation to H. pylori infection and clinicohistological characteristics. *Hepatogastroenterology*. 2002;49(46):1172–6.
- Batley J, Moulding C, Taub R, Murphy W, Stewart T, Potter H, et al. The human c-myc oncogene: structural consequences of translocation into the IgH locus in Burkitt lymphoma. *Cell*. 1983 Oct;34(3):779–87.
- Calcagno D-Q, Leal M-F, Assumpcao P-P, Smith M-A-C, Burbano R-R. MYC and gastric adenocarcinoma carcinogenesis. *World J Gastroenterol*. 2008 Oct;14(39):5962–8.
- Dang C V, O'Donnell KA, Zeller KI, Nguyen T, Osthus RC, Li F. The c-Myc target gene network. *Semin Cancer Biol*. 2006 Aug;16(4):253–64.
- Yasui W, Oue N, Kuniyasu H, Ito R, Tahara E, Yokozaki H. Molecular diagnosis of gastric cancer: present and future. *Gastric cancer Off J Int Gastric Cancer Assoc Japanese Gastric Cancer Assoc*. 2001;4(3):113–21.
- Yasui W, Kudo Y, Semba S, Yokozaki H, Tahara E. Reduced expression of cyclin-dependent kinase inhibitor p27Kip1 is associated with advanced stage and invasiveness of gastric carcinomas. *Jpn J Cancer Res*. 1997 Jul;88(7):625–9.
- Müller W, Noguchi T, Wirtz HC, Hommel G, Gabbert HE. Expression of cell-cycle regulatory proteins cyclin D1, cyclin E, and their inhibitor p21 WAF1/CIP1 in gastric cancer. *J Pathol*. 1999 Oct;189(2):186–93.
- Tsujimoto Y, Finger LR, Yunis J, Nowell PC, Croce CM. Cloning of the chromosome breakpoint of neoplastic B cells with the t(14;18) chromosome translocation. *Science*. 1984 Nov;226(4678):1097–9.
- Tsujimoto Y, Cossman J, Jaffe E, Croce CM. Involvement of the bcl-2 gene in human follicular lymphoma. *Science*. 1985 Jun;228(4706):1440–3.
- Gross A, McDonnell JM, Korsmeyer SJ. BCL-2 family members and the mitochondria in apoptosis. *Genes Dev*. 1999 Aug;13(15):1899–911.
- Vollmers HP, Dämmrich J, Hensel F, Ribbert H, Meyer-Bahlburg A, Ufken-Gaul T, et al. Differential expression of apoptosis receptors on diffuse and intestinal type stomach carcinoma. *Cancer*. 1997 Feb;79(3):433–40.
- Klein R, Vollmers H, MullerHermelink H. Different expression of Bcl-2 in diffuse and intestinal type stomach carcinomas. *Oncol Rep*. 1996;3(5):825–8.
- Yan G-R, Xu S-H, Tan Z-L, Yin X-F, He Q-Y. Proteomics characterization of gastrokine 1-induced growth inhibition of gastric cancer cells. *Proteomics*. 2011 Sep;11(18):3657–64.
- Yoshihara T, Kadota Y, Yoshimura Y, Tatano Y, Takeuchi N, Okitsu H, et al. Proteomic alteration in gastric adenocarcinomas from Japanese patients. *Mol Cancer*. 2006;5(1):75.
- Oien KA, McGregor F, Butler S, Ferrier RK, Downie I, Bryce S, et al. Gastrokine 1 is abundantly and specifically expressed in superficial gastric epithelium, down-regulated in gastric carcinoma, and shows high evolutionary conservation. *J Pathol*. 2004 Jul;203(3):789–97.
- Guo X-Y, Dong L, Qin B, Jiang J, Shi A-M. Decreased expression of gastrokine 1 in gastric mucosa of gastric cancer patients. *World J Gastroenterol*. 2014 Nov;20(44):16702–6.
- Zhang F, Tang JM, Wang L, Shen JY, Zheng L, Wu PP, et al. Detection of  $\beta$ -catenin, gastrokine-2 and embryonic stem cell expressed ras in gastric cancers. *Int J Clin Exp Pathol*. 2010 Nov;3(8):782–91.
- Shimada S, Mimata A, Sekine M, Mogushi K, Akiyama Y, Fukamachi H, et al. Synergistic tumour suppressor activity of E-cadherin and p53 in a conditional mouse model for metastatic diffuse-type gastric cancer. *Gut*. 2012

- Mar;61(3):344–53.
- Tamura G, Yin J, Wang S, Fleisher AS, Zou T, Abraham JM, et al. E-Cadherin gene promoter hypermethylation in primary human gastric carcinomas. *J Natl Cancer Inst.* 2000 Apr;92(7):569–73.
- Mingchao, Devereux TR, Stockton P, Sun K, Sills RC, Clayton N, et al. Loss of E-cadherin expression in gastric intestinal metaplasia and later stage p53 altered expression in gastric carcinogenesis. *Exp Toxicol Pathol Off J Gesellschaft fur Toxikologische Pathol.* 2001 Sep;53(4):237–46.
- Becker KF, Atkinson MJ, Reich U, Becker I, Nekarda H, Siewert JR, et al. E-cadherin gene mutations provide clues to diffuse type gastric carcinomas. *Cancer Res.* 1994 Jul;54(14):3845–52.
- Oda T, Kanai Y, Oyama T, Yoshiura K, Shimoyama Y, Birchmeier W, et al. E-cadherin gene mutations in human gastric carcinoma cell lines. *Proc Natl Acad Sci U S A.* 1994 Mar;91(5):1858–62.
- Guo J, Wang B, Fu Z, Wei J, Lu W. Hypoxic Microenvironment Induces EMT and Upgrades Stem-Like Properties of Gastric Cancer Cells. *Technol Cancer Res Treat.* 2016 Feb;15(1):60–8.
- van Rees BP, Ristimäki A. Cyclooxygenase-2 in carcinogenesis of the gastrointestinal tract. *Scand J Gastroenterol.* 2001 Sep;36(9):897–903.
- Rajnakova A, Mochhala S, Goh PM, Ngoi S. Expression of nitric oxide synthase, cyclooxygenase, and p53 in different stages of human gastric cancer. *Cancer Lett.* 2001 Oct;172(2):177–85.
- van Rees BP, Saukkonen K, Ristimäki A, Polkowski W, Tytgat GNJ, Drillenburger P, et al. Cyclooxygenase-2 expression during carcinogenesis in the human stomach. *J Pathol.* 2002 Feb;196(2):171–9.
- Lim HY, Joo HJ, Choi JH, Yi JW, Yang MS, Cho DY, et al. Increased expression of cyclooxygenase-2 protein in human gastric carcinoma. *Clin cancer Res an Off J Am Assoc Cancer Res.* 2000 Feb;6(2):519–25.
- Saukkonen K, Nieminen O, van Rees B, Vilkkii S, Härkönen M, Juhola M, et al. Expression of cyclooxygenase-2 in dysplasia of the stomach and in intestinal-type gastric adenocarcinoma. *Clin cancer Res an Off J Am Assoc Cancer Res.* 2001 Jul;7(7):1923–31.
- Lee TL, Leung WK, Lau JY, Tong JH, Ng EK, Chan FK, et al. Inverse association between cyclooxygenase-2 overexpression and microsatellite instability in gastric cancer. *Cancer Lett.* 2001 Jul;168(2):133–40.
- Kim H, Lim JW, Kim KH. Helicobacter pylori-induced expression of interleukin-8 and cyclooxygenase-2 in AGS gastric epithelial cells: mediation by nuclear factor-kappaB. *Scand J Gastroenterol.* 2001 Jul;36(7):706–16.
- Song SH, Jong HS, Choi HH, Inoue H, Tanabe T, Kim NK, et al. Transcriptional silencing of Cyclooxygenase-2 by hypermethylation of the 5' CpG island in human gastric carcinoma cells. *Cancer Res.* 2001 Jun;61(11):4628–35.