

Genetic contributions to the pathogenesis of functional dyspepsia

Rajan Singh^a, Uday C Ghoshal^{*b}

^a Department of Physiology and Cell Biology, School of Medicine, University of Nevada Reno, NV, USA

^b Department of Gastroenterology, Sanjay Gandhi Postgraduate Institute of Medical Sciences, Lucknow, India

*Corresponding author e-mail: udayghoshal@gmail.com

ABSTRACT

Functional dyspepsia (FD) is a common gastrointestinal (GI) disorder in the community and in clinical practice. The pathogenesis of FD is largely unknown. However, abnormal gut motility, visceral hypersensitivity, low-grade inflammation, and abnormal brain-gut interaction have been incriminated in its pathogenesis. Accumulating evidence, including the occurrence of these conditions in family members, suggested that functional gastrointestinal disorders (FGIDs), including FD, might be influenced by genetic factors. Several studies have explored the contribution of genetic factors to FD. However, genetic factors contributing to the development of FD remain largely unknown. It appears to be a multi-factorial polygenic disorder. Since gut motility, visceral sensation, and degree of inflammation are all mediated by neuro-peptides, hormones and cytokines and various other proteins, which are transcribed from different genes, alterations in these genes may lead to alterations in these physiological processes. Recent studies showed a host of genetic factors playing important roles in the pathogenesis of FD. We conducted a literature search using the keywords "functional dyspepsia AND gene polymorphisms", "functional dyspepsia AND host genetic factors", and "functional dyspepsia AND gene pathogenesis" to review the major studies on association between FD and gene polymorphisms [*G-protein beta polypeptide-3 (GNβ3)*, *Cholecystokinin-A receptor (CCK-AR)*, *interleukin-17F (IL-17F)*, and *interleukin-10 (IL-10)*].

KEYWORDS: Visceral hypersensitivity, gut-motor dysfunction, low-grade inflammation, functional gastrointestinal disorders; gene polymorphisms

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INTRODUCTION

Splice variant *GNβ3s* is associated with increased intra-cellular signal transduction (Tahara et al; 2008). 825C allele (CC genotype) results in the synthesis of the *GNβ3* wild-type protein predominantly and only minute amounts of the *GNβ3* splice variant (Lin et al; 2001). Also, the homozygous CC genotype is associated with diminished G-protein activity and reduced signal transduction responses (van Lelyveld et al; 2008). Thus, *GNβ3* C825T polymorphism alters intra-cellular signal transduction, which may lead to motor or sensory abnormalities in the GI-tract.

Studies on the association between *GNβ3* C825T and FD have produced inconsistent results. A study from our centre on *GNβ3* C825T polymorphism showed that the homozygous variant TT genotype was more common among patients with FD than HC and conferred up to the two-fold higher risk of FD in reference to homozygous wild CC genotype (Singh et al; 2016). This is perhaps the first study on an Indian population showing an association between *GNβ3* C825T polymorphism and FD. Our findings are consistent with previous studies from Japan, US, Germany, and the Netherlands, showing significant association between this polymorphism and FD (Kim et al; 2012, Oshima et al; 2010, Tahara et al; 2008, van Lelyveld et al; 2008, Camilleri et al; 2006, Chung et al; 2014, Park et al; 2012). Recently, a meta-analysis on eight studies showed a significant association between this polymorphism and FD (Dai et al; 2014). In contrast, studies from Korea showed no association between this polymorphism and FD (Park et al; 2012, Park, 2011). The sample size of patients and controls was small in these studies; hence, the possibility of type II statistical error is quite likely. Inconsistent results have been reported on *GNβ3* C825T polymorphism from the Western countries (Camilleri et al; 2009, van Lelyveld et al; 2008, Camilleri et al; 2006). A study from the Netherlands demonstrated that T allele carriage of *GNβ3* 825C>T polymorphism was associated with dyspepsia (van Lelyveld et al; 2008). A German

study showed that homozygous *GNβ3* 825C status was associated with unexplained dyspepsia (Holtmann et al; 2004). Furthermore, studies from the USA revealed that meal-unrelated dyspepsia was associated with both homozygous *GNβ3* 825TT and CC genotypes (Camilleri et al; 2009, Camilleri et al; 2006). We evaluated this polymorphism in various sub-types of FD [(EPS; epigastric pain syndrome, PDS; post-prandial distress syndrome)]; however, we did not find any association between this polymorphism and various sub-types of FD (It might be due small sample size (Type II error) of EPS and PDS subgroup or due to the fact that patient-reported symptoms might not always correlate with the pathophysiology of FD.) (Singh et al; 2016). Some studies reported a significant association between *GNβ3* C825T polymorphism and sub-types of FD. A study from Japan showed that TT genotype of *GNβ3* C825T was significantly associated with Functional dyspepsia (FD) is a common clinical syndrome characterized by the presence of recurrent or chronic upper abdominal symptoms, such as epigastric pain, early satiety, and fullness, without any structural abnormalities upon upper gastrointestinal (UGI) endoscopy explaining these symptoms (Table 1 and Figure 1) (Drossman, 2006, Stanghellini et al; 2016, Ghoshal et al; 2018, Bhatia et al; 2016, Ghoshal et al; 2011, Shah et al; 2001). FD is a heterogeneous disorder in which different pathophysiological mechanisms underlie specific symptom patterns. Studies have identified several pathophysiological mechanisms, including visceral hypersensitivity, gastric motor dysfunction, and low-grade mucosal inflammation as possible etiological factors (Figure 2) (Li et al; 2013, Takeda et al; 2014, Walker et al; 2014). Visceral hypersensitivity, abnormal gastric motility (impaired gastric accommodation and delayed gastric emptying) and low-grade mucosal inflammation have been considered major pathophysiological mechanisms in FD (Figure 3 and 4) (Mazur et al; 2004, Miwa et al; 2012, Tandon, 2012, Tack et al; 2013, Vanheel; et al, 2013,

Powell et al; 2017). Several studies showed that the patients with FD display exaggerated

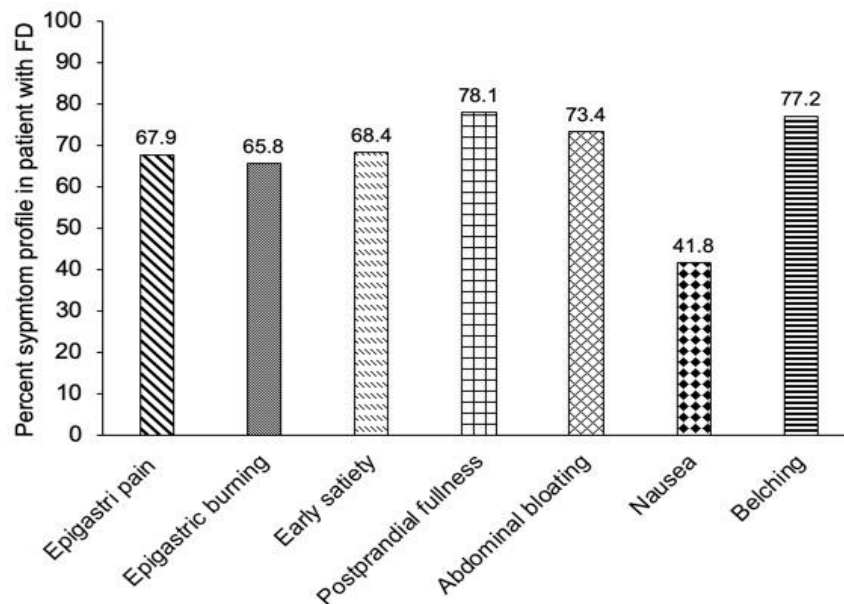


Figure 1. Symptom profile of patients with functional dyspepsia

Table 1: Diagnosis of functional dyspepsia: Rome III criteria

<u>Diagnostic criteria*</u> for functional dyspepsia must include		
1	One or more of	<ul style="list-style-type: none"> Bothersome postprandial fullness Early satiety Epigastric pain Epigastric burning
2	No evidence of structural disease (including at upper endoscopy) that is likely to explain the symptoms	

sensitivity to gastric distension, chemical stimulation and duodenal hypersensitivity to acids (Lee et al; 2010, Vanheel et al; 2013, Vanheel et al; 2014, Mayer et al; 2013). Also, a number of studies implicated inflammatory mechanisms in the pathogenesis of FD (Hughes et al; 2013, Vanheel et al; 2014).

The occurrence of FD might be influenced by multiple genetic factors (Sarnelli et al; 2013). The identification of the genetic factors may improve understanding of the underlying pathophysiological mechanisms. Accumulating evidence, including the occurrence of these conditions in family members, suggested that

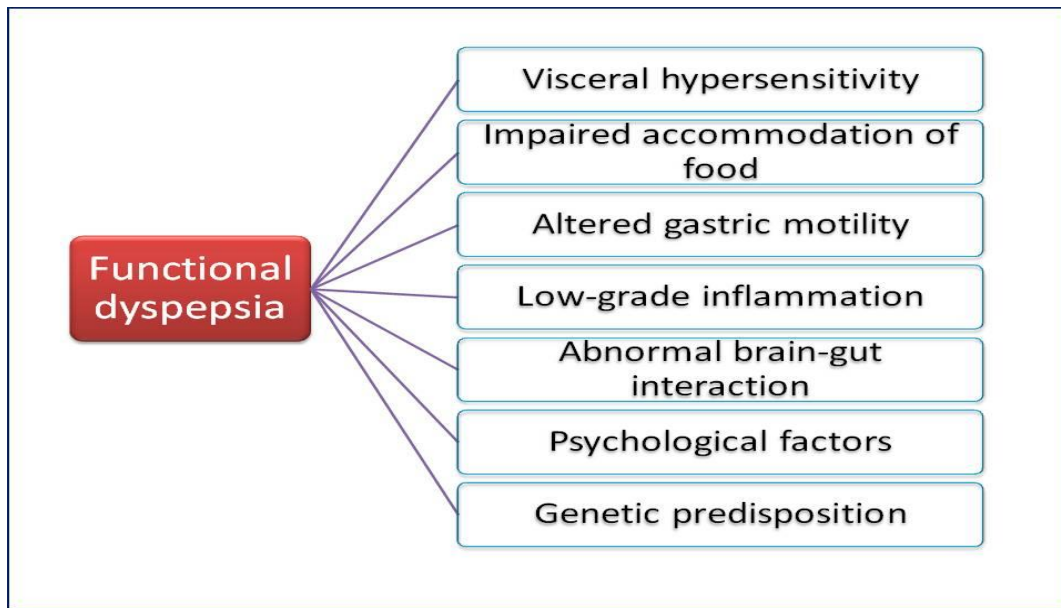


Figure 2. Pathophysiology of functional dyspepsia

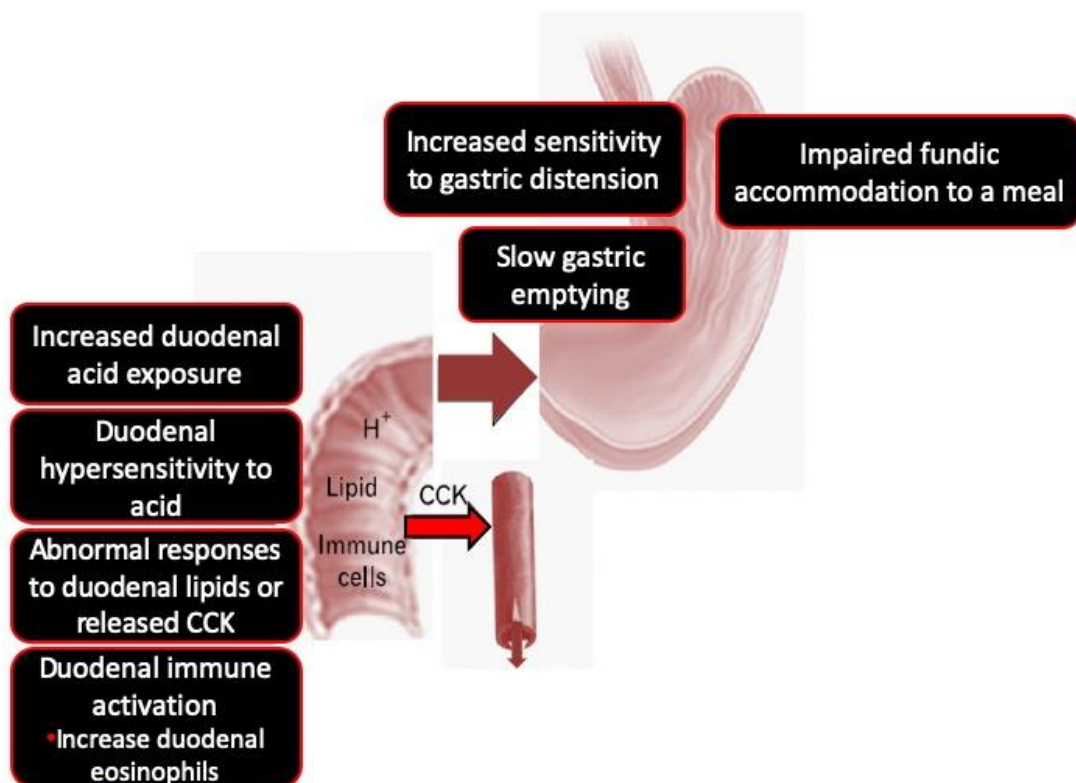


Figure 3. Duodenal visceral hypersensitivity in the pathophysiology of functional dyspepsia

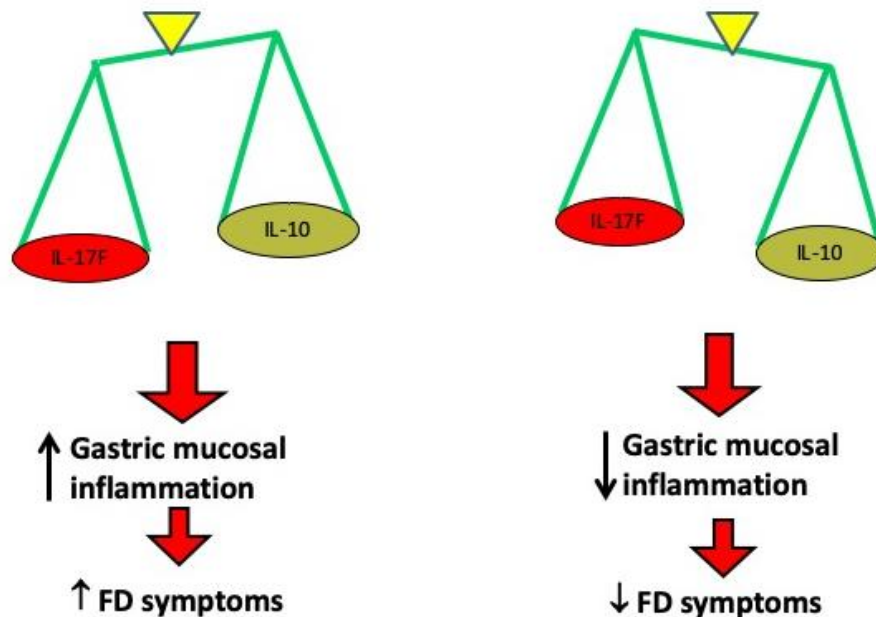


Figure 4. Cytokine gene polymorphism and varying cytokine expression may alter the degree of gastric mucosal inflammation

functional gastrointestinal disorders (FGIDs), including FD, might be influenced by genetic factors (Vanheel et al; 2014, Sarnelli et al, 2013, Adam et al; 2007, Locke et al; 2000, Camilleri et al; 2009). However, genetic factors contributing to the development of FD remain largely unknown. Since, FD might result from altered visceral sensation, gastric motor abnormalities, and low-grade mucosal inflammation in GI, genes influencing it might contribute to the occurrence of FD.

GN β 3 in the pathogenesis of FD

Guanine nucleotide-binding proteins (G-proteins) are essential for stimulus-response in approximately 80% of all known membrane receptors that are linked to intra-cellular signal transduction pathways (Rozengurt et al; 2002, Jankowski et al; 2009). The *G-protein beta polypeptide-3* (GN β 3) gene encodes the G β 3 subunit of G proteins, which are key components of intra-cellular signal transduction (Kim et al;

2012). GN β 3 gene shows common 825C>T polymorphism (rs54443) (Oshima et al; 2010). This substitution 825C>T (TC or TT genotype) is associated with alternative splicing of the gene in exon 9 and the formation of a truncated splice variant GN β 3s, a protein product with 41 amino acids less than in the GN β 3 wild-type protein associated with EPS (Oshima et al; 2010). Another Japanese study showed that CC genotype at this locus was associated with PDS (Shimpuku et al; 2011).

CCK-AR in the pathogenesis of FD

Cholecystokinin (CCK) is an established brain-gut peptide (33-amino acid) that plays an important regulatory role in the GI motility (Pilichiewicz et al; 2008). CCK is released by endocrine I cells within the duodenal and jejunal mucosa in response to various nutrients in the lumen (Pilichiewicz et al; 2008). Biological actions of CCK in the alimentary canal are mediated by CCK-AR (Srivastava et al;

2008). A study showed that fasting and post-fatty meal plasma CCK levels were higher among patients with FD than HC (Pilichiewicz et al; 2008, van Boxel et al; 2014). Intra-venous administration of a CCK-AR antagonist reduced the effects of duodenal lipid on gastric relaxation and gastrointestinal sensations during distension (Maselli et al; 2003). Thus, CCK-AR is involved in the modulation of satiety signal and delayed gastric emptying, which is associated with FD (Fried et al; 2002, Lehmann et al; 2003). *CCK-AR* gene is located on chromosome 4p15. A single nucleotide polymorphism, rs1800857 is located in intron 1 at the intron 1/exon 2 boundary in the vicinity of the mRNA splice acceptor site consensus sequence (Koefoed et al; 2009). Thus, it might result in an alteration of the splicing mechanism, resulting in alteration in the protein (Srivastava et al; 2008, Koefoed et al; 2009). Also, this polymorphism has been widely proposed to play an important role in the regulation of functional mRNA levels of CCK-AR. It is important to study variations in the *CCK-AR* gene, which might alter the response to endogenous CCK and to pharmacologic agents that target these receptors (Srivastava et al; 2008, Chua et al; 2006). There are scant studies on this polymorphism in relation to FD. A study from our centre showed significant an association between *CCK-AR* T/C polymorphism and FD (Singh et al; 2016). A study from Japan reported that this polymorphism was associated with an increased risk of PDS in male subjects (Tahara et al; 2009). Another study from Korea on patients with irritable bowel syndrome-constipation predominant (IBS-C) revealed that *CCK-AR* gene (779T>C) could be functionally important because 779C variant is associated with slower gastric emptying (Park et al; 2010). This observation might suggest that, compared with 779T variant, 779C substitution could result in a greater response to endogenous cholecystokinin, which is known to delay gastric emptying. A study from the US failed to show significant association between *CCK-AR* (779T>C) polymorphism and FD (Camilleri et al; 2009). *CCK-AR* T/C polymorphism

might be associated with reduced levels of CCK-AR. Reduced level of CCK-AR via its ligand CCK might accelerate gastric emptying. Hence, lower FD risk in the presence of this polymorphism is quite expected.

IL-17F in the pathogenesis of FD

Genes encoding cytokines have been reported to be associated with FD (Adam et al; 2007). Moreover, these cytokine genes have polymorphic sites, which are considered to alter gene transcription influencing the degree of the inflammatory response (Adam et al; 2007, Tripathi et al; 2011, Ghoshal U et al; 2013). Allelic variations in their genes may result in high or low production of the cytokines. Therefore, individuals can be classified by their allele status as high, intermediate or low producers of a particular cytokine. A genetic predisposition to produce high or low amounts of a particular cytokine might alter individual susceptibility to FD (Adam et al; 2007, Andersen et al; 2005, Qin et al; 2013). The genetic basis of FD in relation to cytokine production is poorly understood. *IL17F*, a pro-inflammatory molecule, belongs to the IL17 family, which plays a role in coordinating tissue inflammation by inducing the release of other pro-inflammatory and neutrophil-mobilizing cytokines (Gu et al; 2013, Wu et al; 2010). This cytokine is important in neutrophil recruitment and activation causing gastric mucosal inflammation (Arisawa et al; 2007). Since gastritis is associated with symptoms of FD, it is important to study the role of immune related gene polymorphism, such as that of *IL17F*, in the pathophysiology of FD. Cytogenetic location of *IL17F* is on 6p12 (Paradowska-Gorycka et al; 2010). This gene exhibits common single nucleotide polymorphisms (SNPs) rs2397084: A/G and rs763780: T/C. The functional role of *IL17F* (rs2397084) polymorphism is not well known. However, some studies suggest that it may alter *IL17F* expression (Paradowska-Gorycka et al; 2010). *IL17F* (rs763780) polymorphism causes His-to-Arg substitution at amino acid 161 (H161R), which leads

to the reduction of *IL17-F* cytokines level (Wu et al; 2010).

In a study from our centre, we found that GG (variant) genotype of *IL17F* (rs2397084) was associated with FD as compared to the presence of AA (wild) genotype (Singh et al; 2017). Such a high frequency of variant genotype in FD suggests that high production of IL17F (a pro-inflammatory cytokine) may have some risk for FD. Such genetic predisposition to higher pro-inflammatory cytokine production among patients with FD might mean that exaggerated inflammatory response may predispose to FD. This is perhaps the first study on *IL17F* (rs2397084) polymorphism in FD. This SNP has been studied in other inflammatory disorders such as lung cancer and rheumatoid arthritis; however, no significant association was found in those diseases (Paradowska-Gorycka et al; 2010, Kaabachi et al; 2014). Our study supported the recently proposed hypothesis that FD may be related to mucosal low-grade inflammation (Barbara et al; 2016, Enck et al; 2017). Thus, a genetic predisposition to produce high amounts of pro-inflammatory cytokines might alter individual susceptibility to FD. Another gene polymorphism we studied, *IL17F* (rs763780), was comparable among patients with FD and with HS (Singh et al; 2017). This polymorphism causes His-to-Arg substitution at amino acid 161 (H161R), which leads to reduction of *IL17F* cytokines level (Arisawa et al; 2007). In contrast, one Japanese study showed that the *T* allele of *IL17F* (rs763780) was significantly associated with EPS, a subgroup of FD (Arisawa et al; 2007). Another study from Japan showed that *IL17F* (rs763780) polymorphism was comparable among patients with duodenal ulcer disease and HS (Hayashi et al; 2012). Also, some studies reported an association between *IL17F* (rs763780) and ulcerative colitis, another inflammatory gastrointestinal condition (Li et al; 2014).

IL10 in the pathogenesis of FD

IL10 is an immune-regulatory cytokine that inhibits cell-mediated immune response (Olszak et al;

2014). It has been shown to exert potent anti-inflammatory activity by down regulating pro-inflammatory cytokines such as *TNF- α* , *IL1*, *IL12*, and chemokines (Olszak et al; 2014, Bodger et al; 2001). A genetic predisposition to produce low levels of anti-inflammatory cytokines may result in an inability to control the inflammatory response, which may be associated with the development of some types of FGIDs including FD (Adam et al; 2007, Qin et al; 2013).

IL10 gene is located on chromosome 1q31-32 and consists of five exons and four introns (Rad et al; 2004). Two commonly studied polymorphisms in the *IL10* gene are rs1800896: G/A and rs1800871: C/T (Rad et al; 2004, Zhao et al; 2014). *IL10* -819 C/T and -592 A/C showed 100% linkage disequilibrium in their genotype distribution (Rad et al; 2004). It has been reported that *IL10* SNPs markedly influenced mucosal *IL10* expression in the course of chronic *H. pylori* infection: GCC haplotype carriers (*IL10* -1082G; -819C; -592C) were associated with high and ATA carriers with low *IL10* expression (Rad et al; 2004).

In a study from our centre, we showed that *IL10* (rs1800896) polymorphism was comparable among patients with FD and HS (Singh et al; 2017). However, TT (under-producers) genotype of *IL10* (rs1800871) was more common among patients than HS. Such a high frequency of the under-producer genotype in FD suggests that lowers the production of *IL10* (an anti-inflammatory cytokine) may have some risk for FD. Several studies have shown that serum *IL10* levels are significantly lower among patients with IBS (another common FGID) than HS, suggesting that altered *IL10* levels may be involved in the pathogenesis of FGIDs (Qin et al; 2013, Zhao et al; 2014, Bashashati et al; 2014). Also, it has been reported that *IL10* polymorphisms markedly influenced mucosal *IL10* expression in the course of chronic *H. pylori* infection: GCC haplotype carriers (*IL10* -1082G; -819C; -592C) were associated with high and ATA carriers with low *IL10* expression (Rad et al; 2004). A study by Wang BM et al showed that *IL10* genotypes, -819 T/T and -592 A/A were associated

with diarrhea-predominant irritable bowel syndrome (D-IBS) (Wang et al; 2006). In another study, low prevalence of high-producer genotype in patients with IBS suggested that high levels of *IL10* might have some protective role and low producers of this cytokine might be more likely to develop IBS (Qin et al; 2013, Gonsalkorale et al; 2003, Bashashati et al; 2012). A genetic predisposition to produce low levels of anti-inflammatory cytokines could mean a compromised control of the inflammatory response, which may lead to continuing problems.

Conclusion and future directions

Several studies showed *GNβ3* C825T polymorphism was associated with FD.

Homozygous CC genotype of *CCK-AR* T/C polymorphism (associated with lower expression of *CCKAR*) is protective against FD. The importance of visceral hypersensitivity and gut motility related gene polymorphisms in FD has been emphasized. In addition, these results provide some support to the concept that 'altered gastrointestinal motor and sensory function' may contribute to the pathogenesis of this condition. Polymorphisms in immune related genes are involved in the development of FD. *IL17F* (rs2397084) and *IL10* (rs1800871) are associated with a higher risk for FD. These studies provided evidence that the genetic polymorphisms in genes associated with inflammation or immune response may be involved in the development of FD (Figure 5).

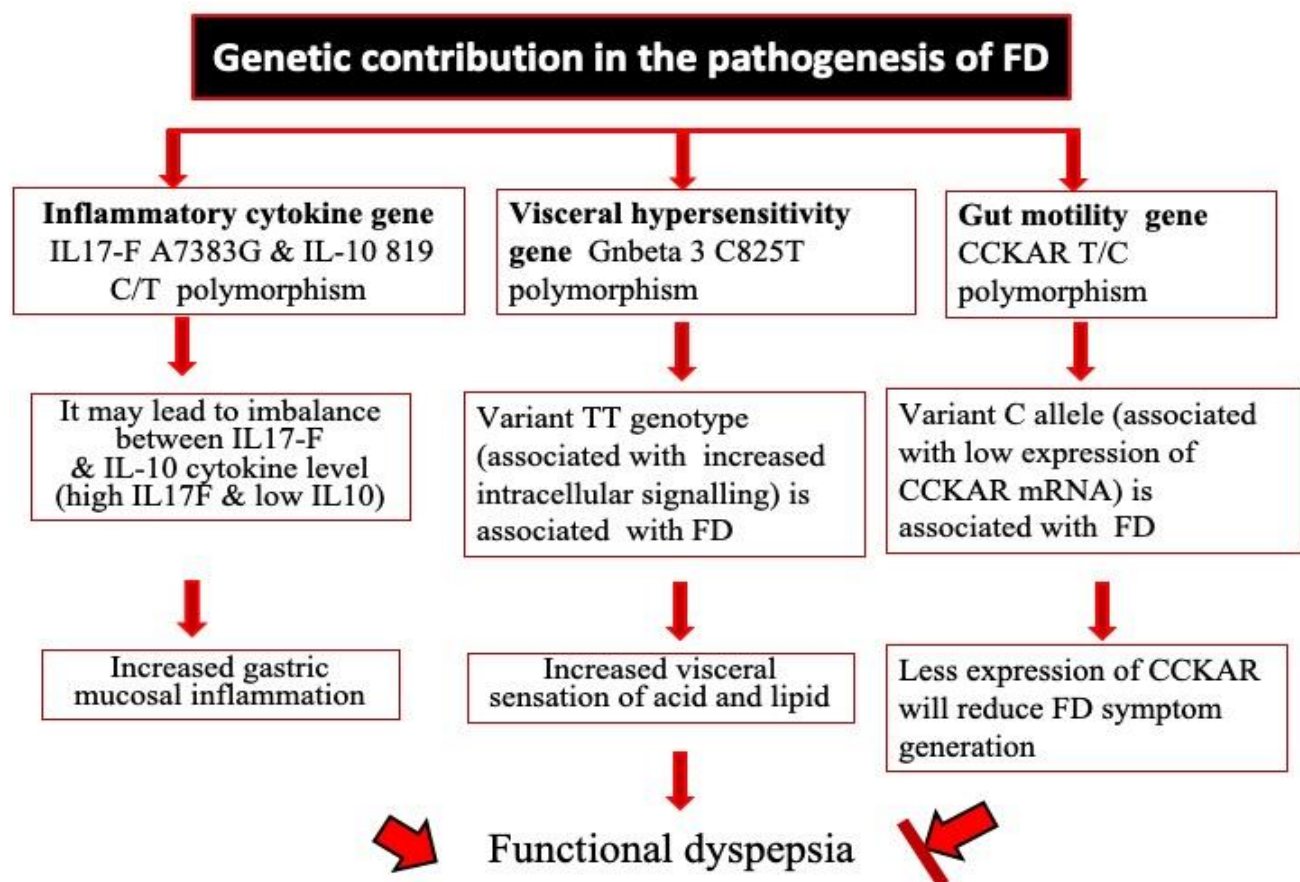


Figure 5. Summary of genetic polymorphism study

In addition, these studies provided some support to the concept that 'low-grade inflammation' might contribute to the pathogenesis of this condition. Future studies should replicate these association studies in different populations and also explain the molecular mechanisms of pathogenesis of the condition. Animal studies related to an individual gene knock down/knock out/knock in warrant further studies to explore the exact molecular mechanisms in sub-types of FD and disease progression.

Conflict of interest statement

The authors declared to have no competing or conflict of interest.

Authors' contributions

RS and UCG conceived, designed and drafted the manuscript.

REFERENCES

- Adam B, Liebrechts T, Holtmann G. Mechanisms of disease: genetics of functional gastrointestinal disorders-searching the genes that matter. *Nat Clin Pract Gastroenterol Hepatol* 2007; 4:102-10.
- Andersen LP, Holck S, Janulaityte-Gunther D, Kupcinskas L, Kiudelis G, Jonaitis L et al. Gastric inflammatory markers and interleukins in patients with functional dyspepsia, with and without *Helicobacter pylori* infection. *FEMS Immunol Med Microbiol* 2005; 44:233-8.
- Arisawa T, Tahara T, Shibata T, Nagasaka M, Nakamura M, Kamiya Y et al. Genetic polymorphisms of molecules associated with inflammation and immune response in Japanese subjects with functional dyspepsia. *Int J Mol Med* 2007; 20:717-23.
- Barbara G, Feinle-Bisset C, Ghoshal UC, et al. The Intestinal Microenvironment and Functional Gastrointestinal Disorders. *Gastroenterology*. 2016 Feb 18. pii: S0016-5085(16)00219-5.
- Bashashati M, Rezaei N, Bashashati H, Shafieyoun A, Daryani NE, Sharkey KA et al. Cytokine gene polymorphisms are associated with irritable bowel syndrome: a systematic review and meta-analysis. *Neurogastroenterol Motil* 2012; 24:1102-e566.
- Bashashati M, Rezaei N, Shafieyoun A, McKernan DP, Chang L, Ohman L, Quigley EM et al. Cytokine imbalance in irritable bowel syndrome: a systematic review and meta-analysis. *Neurogastroenterol Motil* 2014; 26:1036-48.
- Bhatia V, Deswal S, Seth S, Kapoor A, Sibal A, Gopalan S. Prevalence of functional gastrointestinal disorders among adolescents in Delhi based on Rome III criteria: A school-based survey. *Indian J Gastroenterol*. 2016 Jul;35(4):294-8.
- Bodger K, Bromelow K, Wyatt JI, Heatley RV. Interleukin 10 in *Helicobacter pylori* associated gastritis: immunohistochemical localisation and in vitro effects on cytokine secretion. *J Clin Pathol* 2001; 54:285-92.
- Camilleri CE, Carlson PJ, Camilleri M, Castillo EJ, Locke GR, 3rd, Geno DM et al. A study of candidate genotypes associated with dyspepsia in a U.S. community. *Am J Gastroenterol* 2006; 101:581-92.
- Camilleri M, Zinsmeister AR. Candidate genes and functional dyspepsia. *Neurogastroenterol Motil* 2009; 21:94.
- Chua AS, Keeling PW, Dinan TG. Role of cholecystokinin and central serotonergic receptors in functional dyspepsia. *World J Gastroenterol* 2006; 12:1329-35.
- Chung HA, Lee SY, Lee HJ, Kim JH, Sung IK, Shim CS et al. G protein beta3 subunit polymorphism and long-term prognosis of functional dyspepsia. *Gut Liver* 2014; 8:271-6.
- Dai F, Liu Y, Shi H, Ge S, Song J, Dong L et al. Association of genetic variants in GNbeta3 with functional dyspepsia: a meta-analysis. *Dig Dis Sci* 2014; 59:1823-30.
- Drossman DA. The functional gastrointestinal disorders and the Rome III process. *Gastroenterology* 2006; 130: 1377-90.
- Enck P, Azpiroz F, Boeckxstaens G et al. Functional dyspepsia. *Nat Rev Dis Primers*. 2017 Nov 3; 3:17081.
- Fried M, Feinle C. The role of fat and cholecystokinin in functional dyspepsia. *Gut* 2002;51 Suppl 1: i54-7.
- Ghoshal U, Kumar S, Jaiswal V, Tripathi S, Mittal B, Ghoshal UC. Association of microsomal epoxide hydrolase exon 3 Tyr113His and exon 4 His139Arg polymorphisms with gastric cancer in India. *Indian J Gastroenterol*. 2013 Jul;32(4):246-52.
- Ghoshal UC, Sachdeva S, Pratap N, Verma A, Karyampudi A, Misra A et al. Indian consensus on chronic constipation in adults: A joint position statement of the Indian Motility and Functional Diseases Association and the Indian Society of Gastroenterology. *Indian J Gastroenterol*. 2018 Nov;37(6):526-544.
- Ghoshal UC, Singh R, Chang FY, Hou X, Wong BC, Kachintorn U et al. Epidemiology of uninvestigated and functional dyspepsia in Asia: facts and fiction. *J Neurogastroenterol Motil*. 2011 Jul;17(3):235-44.
- Gonsalkorale WM, Perrey C, Pravica V, Whorwell PJ, Hutchinson IV. Interleukin 10 genotypes in irritable bowel syndrome: evidence for an inflammatory component? *Gut* 2003; 52:91-3.
- Gu C, Wu L, Li X. IL-17 family: cytokines, receptors and signaling. *Cytokine* 2013; 64:477-85.
- Hayashi R, Tahara T, Shiroeda H, Matsue Y, Minato T, Nomura T et al. Association of genetic polymorphisms in IL17A

- and IL17F with gastro-duodenal diseases. *J Gastrointest Liver Dis* 2012; 21:243-9.
- Holtmann G, Siffert W, Haag S, Mueller N, Langkafel M, Senf W et al. G-protein beta 3 subunit 825 CC genotype is associated with unexplained (functional) dyspepsia. *Gastroenterology* 2004; 126:971-9.
- Hughes PA, Harrington AM, Castro J, Liebrechts T, Adam B, Grasby DJ et al. Sensory neuro-immune interactions differ between irritable bowel syndrome subtypes. *Gut* 2013; 62:1456-65.
- Jankowski JA, Talley NJ. Dissecting GI phenotype-genotype relationships in GERD and dyspepsia: an SNP here and an SNP there! *Am J Gastroenterol* 2009; 104:286-8.
- Kaabachi W, ben Amor A, Kaabachi S, Rafrafi A, Tizaoui K, Hamzaoui K. Interleukin-17A and -17F genes polymorphisms in lung cancer. *Cytokine* 2014; 66:23-9.
- Kim HG, Lee KJ, Lim SG, Jung JY, Cho SW. G-Protein Beta3 Subunit C825T Polymorphism in Patients With Overlap Syndrome of Functional Dyspepsia and Irritable Bowel Syndrome. *J Neurogastroenterol Motil* 2012; 18:205-10.
- Koefoed P, Hansen TV, Woldbye DP, Werge T, Mors O, Hansen T et al. An intron 1 polymorphism in the cholecystokinin-A receptor gene associated with schizophrenia in males. *Acta Psychiatr Scand* 2009; 120:281-7.
- Lee KJ, Tack J. Duodenal implications in the pathophysiology of functional dyspepsia. *J Neurogastroenterol Motil* 2010; 16:251-7.
- Lehmann F, Hildebrand P, Beglinger C. New molecular targets for treatment of peptic ulcer disease. *Drugs* 2003; 63:1785-97.
- Li J, Tian H, Jiang HJ, Han B. Interleukin-17 SNPs and serum levels increase ulcerative colitis risk: a meta-analysis. *World J Gastroenterol* 2014; 20:15899-909.
- Li X, Cao Y, Wong RK, Ho KY, Wilder-Smith CH. Visceral and somatic sensory function in functional dyspepsia. *Neurogastroenterol Motil* 2013; 25:246-53, e165.
- Lin CN, Tsai SJ, Hong CJ. Association analysis of a functional G protein beta3 subunit gene polymorphism (C825T) in mood disorders. *Neuropsychobiology* 2001; 44:118-21.
- Locke GR, 3rd, Zinsmeister AR, Talley NJ, Fett SL, Melton LJ, 3rd. Familial association in adults with functional gastrointestinal disorders. *Mayo Clin Proc* 2000; 75:907-12.
- Maselli MA, Mennuni L. CCK1 receptor antagonist, dexloxiglumide: effects on human isolated gallbladder. Potential clinical applications. *Minerva Gastroenterol Dietol* 2003; 49:211-6.
- Mayer EA, Tillisch K, Ellingson BM. Dyspepsia: Structural changes in functional gastrointestinal disorders. *Nat Rev Gastroenterol Hepatol* 2013; 10:200-2.
- Mazur M, Furgala A, Thor PJ. [Visceral sensitivity disturbances in the pathogenesis of functional gastrointestinal disorders]. *Folia Med Cracov* 2004; 45:33-49.
- Miwa H. [Functional dyspepsia from pathogenic perspective]. *Nihon Shokakibyō Gakkai Zasshi* 2012; 109: 1683-96.
- Olszak T, Neves JF, Dowds CM, Baker K, Glickman J, Davidson NO et al. Protective mucosal immunity mediated by epithelial CD1d and IL-10. *Nature* 2014; 509:497-502.
- Oshima T, Nakajima S, Yokoyama T, Toyoshima F, Sakurai J, Tanaka J et al. The G-protein beta3 subunit 825 TT genotype is associated with epigastric pain syndrome-like dyspepsia. *BMC Med Genet* 2010; 11:13.
- Paradowska-Gorycka A, Wojtecka-Lukasik E, Trefler J, Wojciechowska B, Lacki JK, Maslinski S. Association between IL-17F gene polymorphisms and susceptibility to and severity of rheumatoid arthritis (RA). *Scand J Immunol* 2010; 72:134-41.
- Park CS, Uhm JH. Polymorphisms of the Serotonin Transporter Gene and G-Protein beta3 Subunit Gene in Korean Children with Irritable Bowel Syndrome and Functional Dyspepsia. *Gut Liver* 2012; 6:223-8.
- Park H. Functional gastrointestinal disorders and overlap syndrome in Korea. *J Gastroenterol Hepatol* 2011; 26 Suppl 3:12-4.
- Park MI. Is There Enough Evidence for the Association of GNbeta3 C825T Polymorphism With Functional Dyspepsia and Irritable Bowel Syndrome? *J Neurogastroenterol Motil* 2012; 18:348-9.
- Park SY, Rew JS, Lee SM, Ki HS, Lee KR, Cheo JH et al. Association of CCK(1) Receptor Gene Polymorphisms and Irritable Bowel Syndrome in Korean. *J Neurogastroenterol Motil* 2010; 16:71-6.
- Pilichiewicz AN, Feltrin KL, Horowitz M, Holtmann G, Wishart JM, Jones KL et al. Functional dyspepsia is associated with a greater symptomatic response to fat but not carbohydrate, increased fasting and postprandial CCK, and diminished PYY. *Am J Gastroenterol* 2008; 103:2613-23.
- Powell N, Walker MM, Talley NJ. The mucosal immune system: master regulator of bidirectional gut-brain communications. *Nat Rev Gastroenterol Hepatol*. 2017 Mar;14(3):143-159.
- Qin SY, Jiang HX, Lu DH, Zhou Y. Association of interleukin-10 polymorphisms with risk of irritable bowel syndrome: a meta-analysis. *World J Gastroenterol* 2013; 19: 9472-80.
- Rad R, Dossumbekova A, Neu B, Lang R, Bauer S, Saur D et al. Cytokine gene polymorphisms influence mucosal cytokine expression, gastric inflammation, and host specific colonisation during *Helicobacter pylori* infection. *Gut* 2004; 53:1082-9.
- Rozengurt E, Guha S, Sinnott-Smith J. Gastrointestinal peptide signalling in health and disease. *Eur J Surg Suppl* 2002:23-38.
- Sarnelli G, D'Alessandro A, Pesce M, Palumbo I, Cuomo R. Genetic contribution to motility disorders of the upper gastrointestinal tract. *World J Gastrointest Pathophysiol* 2013; 4: 65-73.

- Shah SS, Bhatia SJ, Mistry FP. Epidemiology of dyspepsia in the general population in Mumbai. *Indian J Gastroenterol*. 2001 May-Jun;20(3):103-6.
- Shimpuku M, Futagami S, Kawagoe T, Nagoya H, Shindo T, Horie A et al. G-protein beta3 subunit 825CC genotype is associated with postprandial distress syndrome with impaired gastric emptying and with the feeling of hunger in Japanese. *Neurogastroenterol Motil* 2011; 23:1073-80.
- Singh R, Ghoshal UC, Kumar S et al. Genetic variants of immune-related genes IL17F and IL10 are associated with functional dyspepsia: A case-control study. *Indian J Gastroenterol*. 2017 Sep;36(5):343-352.
- Singh R, Mittal B, Ghoshal UC. Functional dyspepsia is associated with GN β 3 C825T and CCK-AR T/C polymorphism. *Eur J Gastroenterol Hepatol*. 2016 Feb;28(2):226-32.
- Srivastava A, Pandey SN, Dixit M, Choudhuri G, Mittal B. Cholecystokinin receptor A gene polymorphism in gallstone disease and gallbladder cancer. *J Gastroenterol Hepatol* 2008; 23:970-5.
- Stanghellini V, Chan FK, Hasler WL et al. Gastrointestinal Disorders. *Gastroenterology*. 2016 May;150(6):1380-92.
- Tack J, Talley NJ. Functional dyspepsia--symptoms, definitions and validity of the Rome III criteria. *Nat Rev Gastroenterol Hepatol*. 2013 Mar;10(3):134-41.
- Tahara T, Arisawa T, Shibata T, Nakamura M, Wang F, Yoshioka D et al. 779 TC of CCK-1 intron 1 is associated with postprandial syndrome (PDS) in Japanese male subjects. *Hepatogastroenterology* 2009; 56:1245-8.
- Tahara T, Arisawa T, Shibata T, Wang F, Nakamura M, Sakata M et al. Homozygous 825T allele of the GNB3 protein influences the susceptibility of Japanese to dyspepsia. *Dig Dis Sci* 2008; 53:642-6.
- Takeda H. [Recent mechanistic insights into the pathogenesis of functional dyspepsia: focusing on interoceptive system]. *Nihon Shokakibyō Gakkai Zasshi* 2014; 111:1058-70.
- Tandon RK. Etiopathogenesis of functional dyspepsia. *J Assoc Physicians India* 2012;60 Suppl:18-20.
- Tripathi S, Ghoshal U, Mittal B, Chourasia D, Kumar S, Ghoshal UC. Association between gastric mucosal glutathione-S-transferase activity, glutathione-S-transferase gene polymorphisms and *Helicobacter pylori* infection in gastric cancer. *Indian J Gastroenterol*. 2011 Dec; 30(6):257-63.
- van Boxel OS, ter Linde JJ, Oors J, Otto B, Weusten BL, Feinle-Bisset C et al. Functional dyspepsia patients have lower mucosal cholecystokinin concentrations in response to duodenal lipid. *Eur J Gastroenterol Hepatol* 2014; 26:205-12.
- van Lelyveld N, Linde JT, Schipper M, Samsom M. Candidate genotypes associated with functional dyspepsia. *Neurogastroenterol Motil* 2008; 20:767-73.
- Vanheel H, Farré R. Changes in gastrointestinal tract function and structure in functional dyspepsia. *Nat Rev Gastroenterol Hepatol*. 2013 Mar;10 (3):142-9.
- Vanheel H, Vicario M, Vanuytsel T, Van Oudenhove L, Martinez C, Keita AV, Pardon N et al. Impaired duodenal mucosal integrity and low-grade inflammation in functional dyspepsia. *Gut* 2014; 63:262-71.
- Walker MM, Aggarwal KR, Shim LS, Bassan M, Kalantar JS, Weltman MD et al. Duodenal eosinophilia and early satiety in functional dyspepsia: confirmation of a positive association in an Australian cohort. *J Gastroenterol Hepatol* 2014; 29: 474-9.
- Wang BM, Jiang XZ, Yang YL, Liu WT, Cao XC, Zhao XZ. [A study of interleukin-10 gene polymorphism in irritable bowel syndrome]. *Zhonghua Nei Ke Za Zhi* 2006; 45:289-92.
- Wu X, Zeng Z, Chen B, Yu J, Xue L, Hao Y et al. Association between polymorphisms in interleukin-17A and interleukin-17F genes and risks of gastric cancer. *Int J Cancer* 2010; 127:86-92.
- Zhao L, Song W, Zhu P, Zhang Y, Bu P. [A correlation study between diarrhea-predominant irritable bowel syndrome complicated functional dyspepsia patients of Gan-stagnation Pi-deficiency syndrome and gastrointestinal hormones]. *Zhongguo Zhong Xi Yi Jie He Za Zhi* 2014; 34:1168-72.